

# Μικροσκοπία Φθορισμού: Τεχνολογία και Εφαρμογές

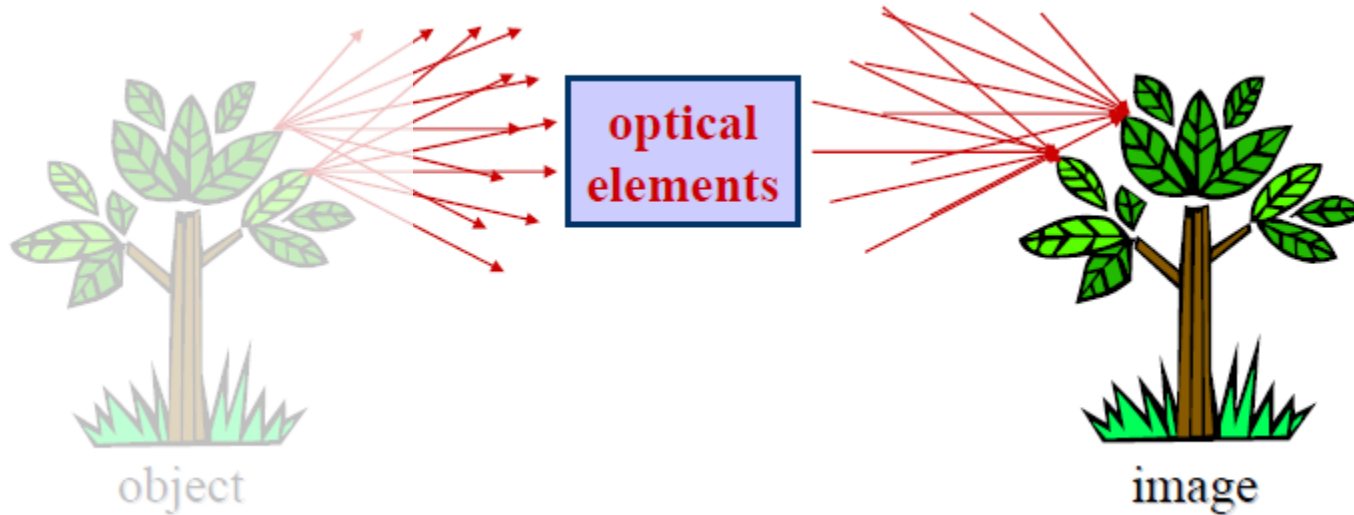
Δημήτριος Τζεράνης, Ph.D.

Εμβιομηχανική και Βιοϊατρική Τεχνολογία  
Τμήμα Μηχανολόγων Μηχανικών | Ε.Μ.Π.

Χειμερινό Εξάμηνο 2015

# The Three Components of Imaging

- Three major components



## Emission generation

- Fluorescence

## Optical system

- Microscope
- Endoscope

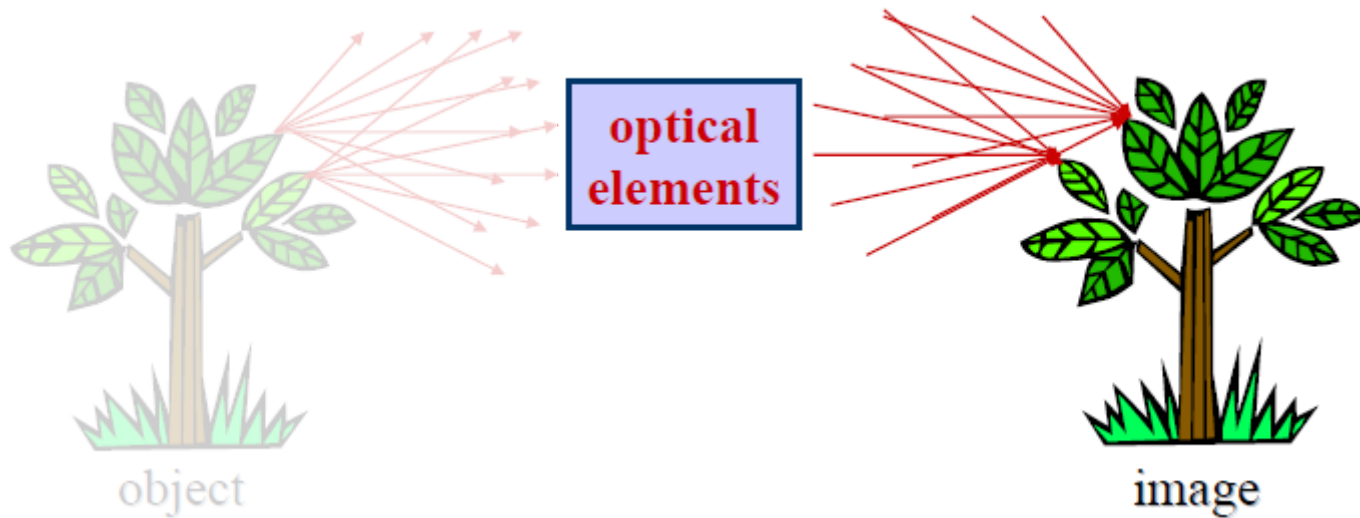
## Emission detection

- Detector & sampling

# Presentation Overview

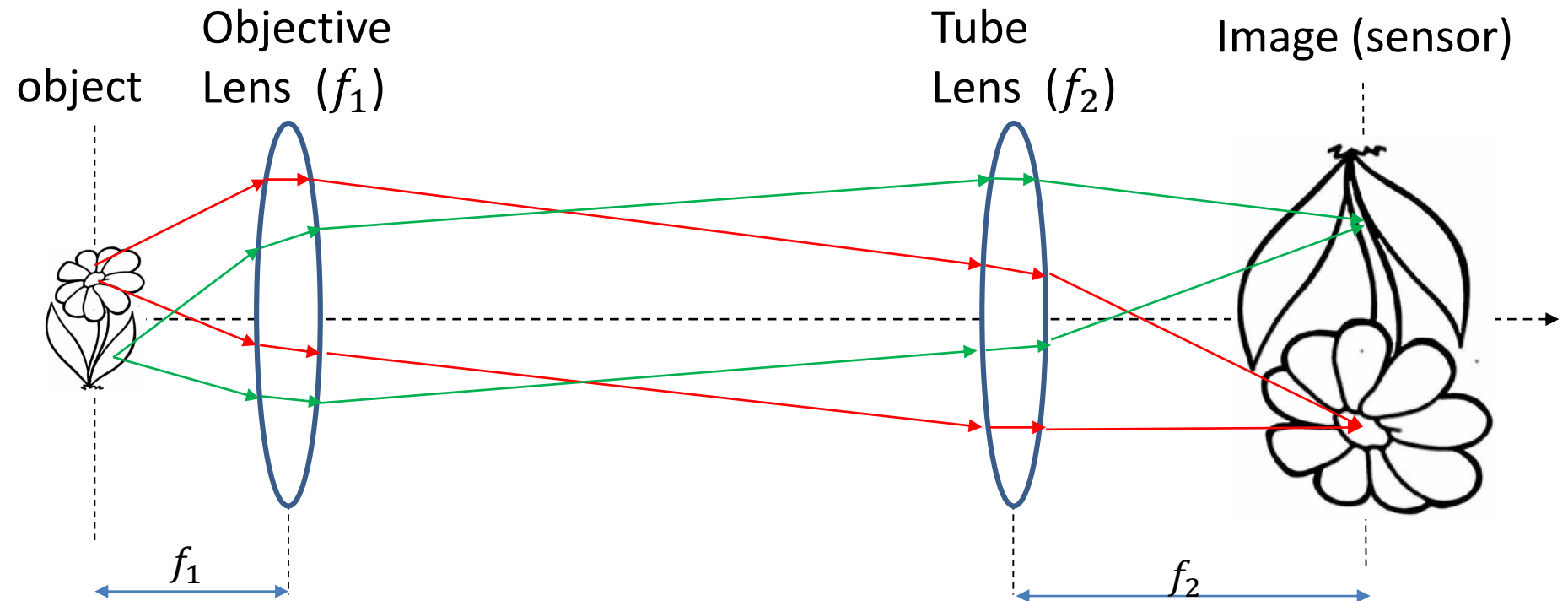
- Microscopy basics
- Standard fluorescence microscopy
- State-of-the-art and Applications of fluorescence microscopy

# Microscopy Basics



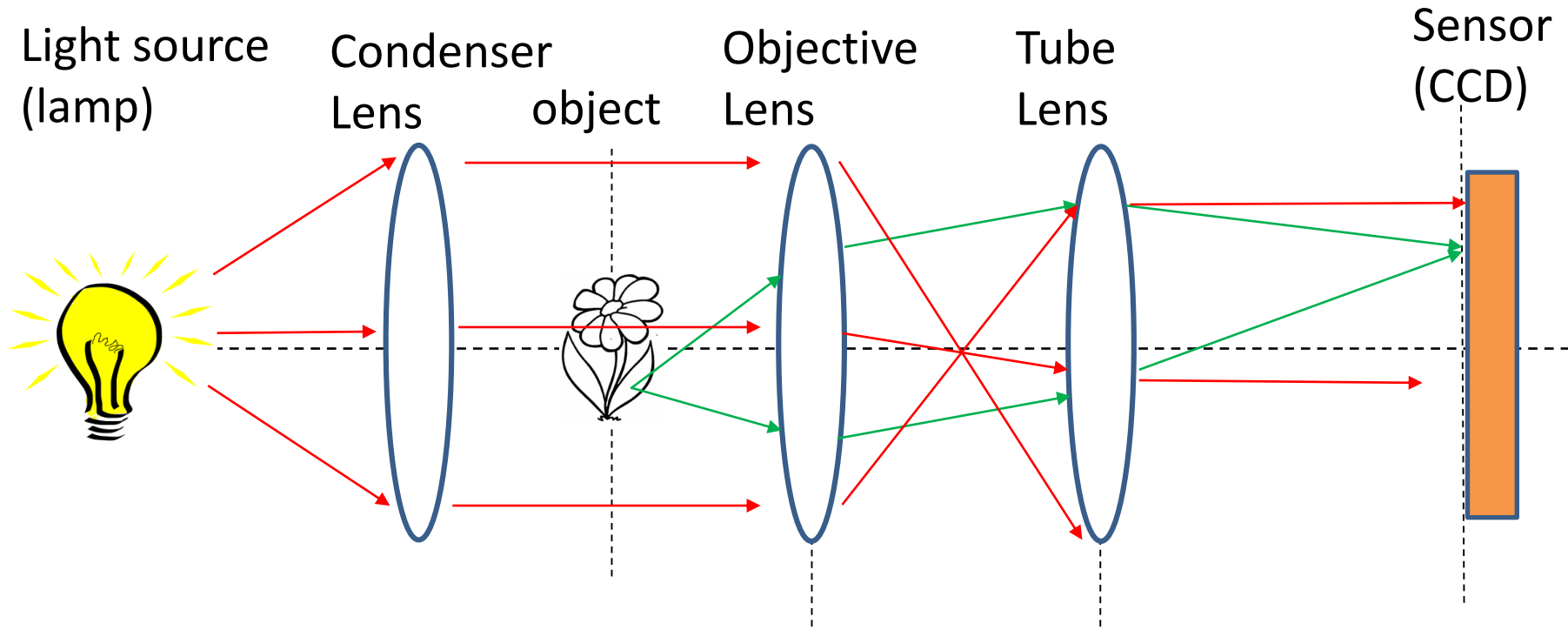
# Microscope Principle

- Utilize an optical system to magnify objects
  - Simplest system: 2 lenses
  - Magnification:  $M = \frac{f_2}{f_1}$

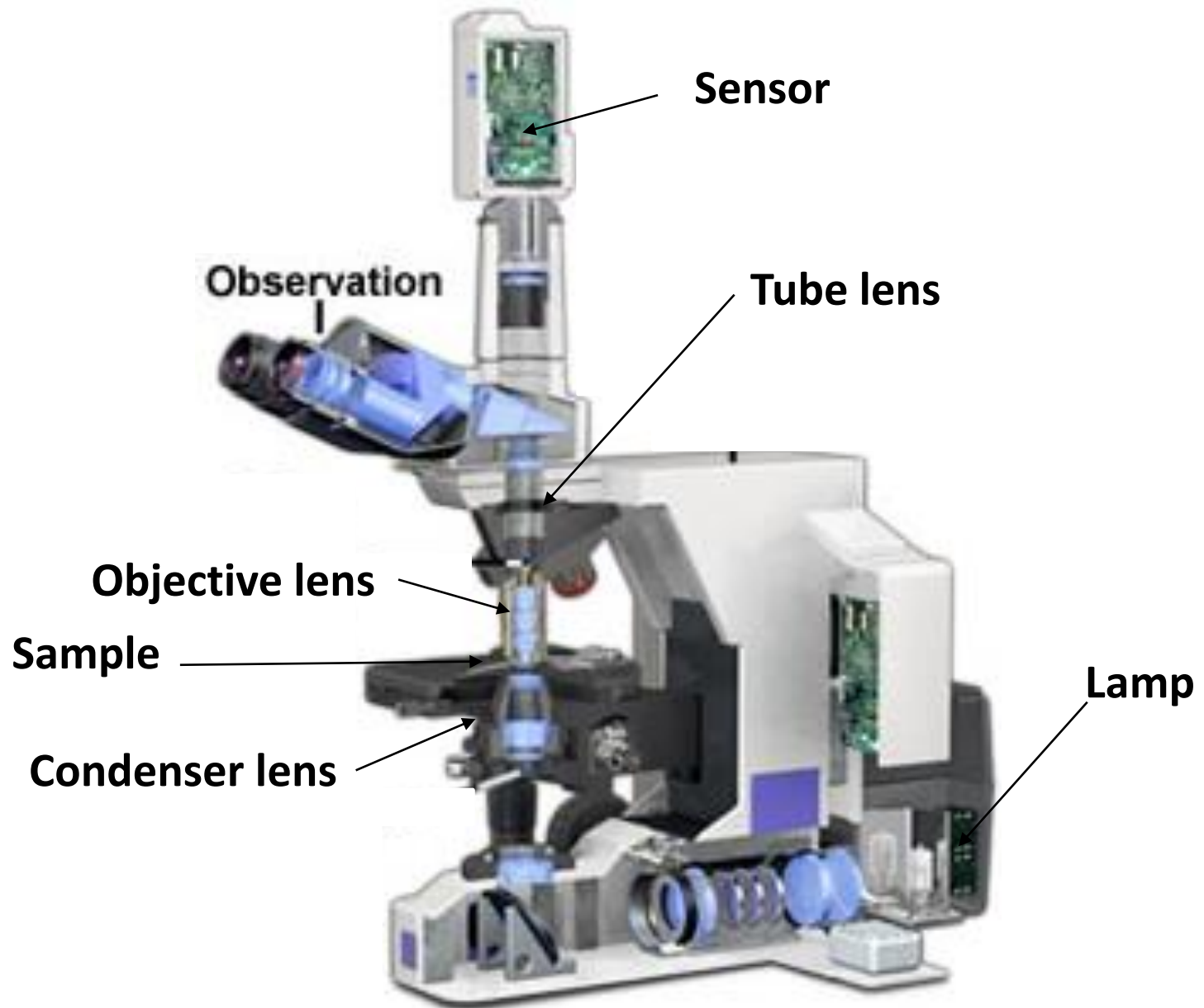


# Microscope Principle

- Need a light source (illumination) to generate useful signal (emission) from sample
  - Some illumination is also detected  $\rightarrow$  reduce SNR
  - Ideally would like to detect only emission

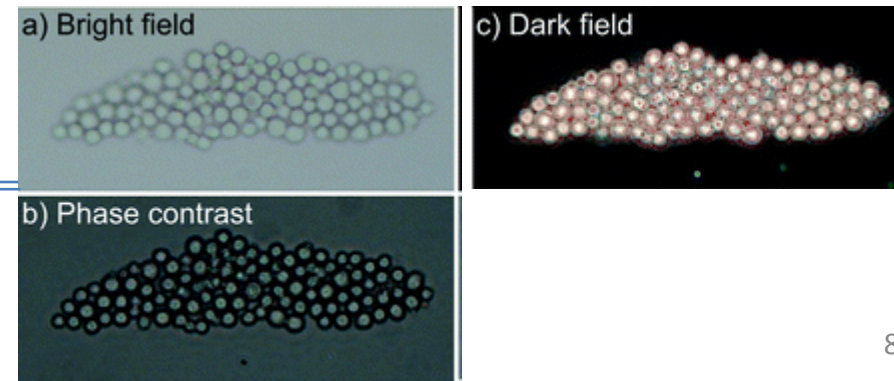
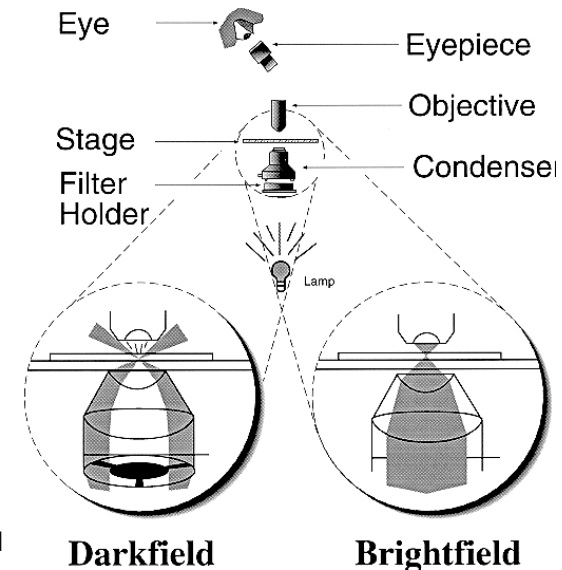
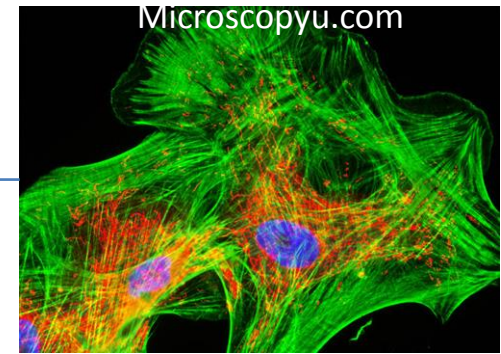


# Microscope Uncovered



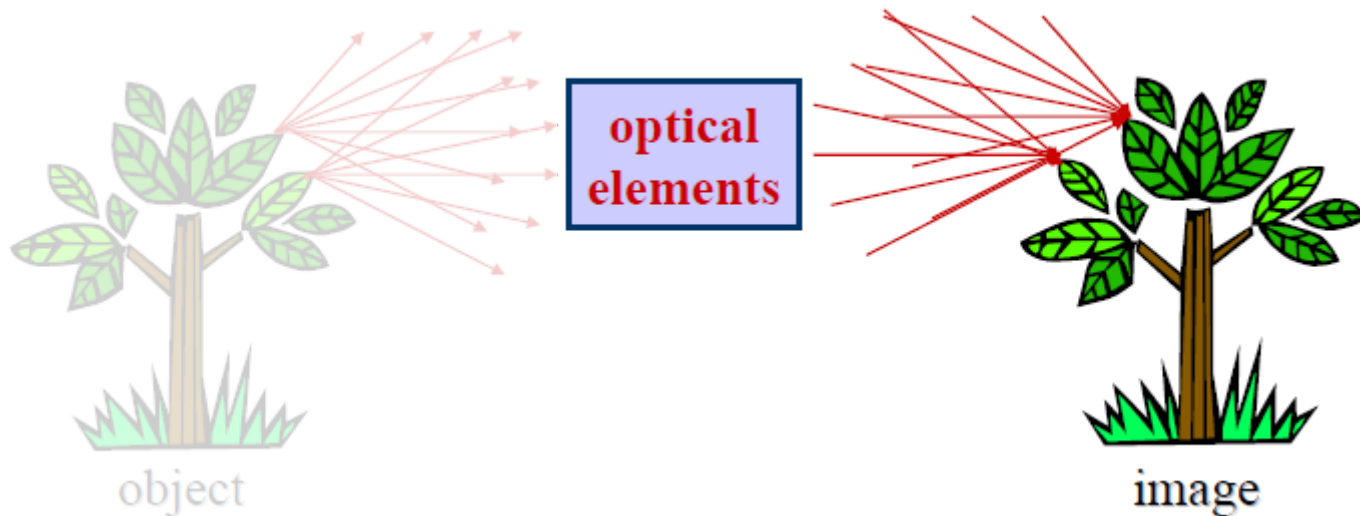
# Microscopy Modes Utilized in Biology & Medicine

- Modes: ways to generate & detect emission in sample
- Fluorescence microscopy
  - Focus of this presentation
- Brightfield, darkfield microscopy
  - Standard mode
- Phase microscopy
  - Image very thin & transparent samples





# Standard Fluorescence Microscopy

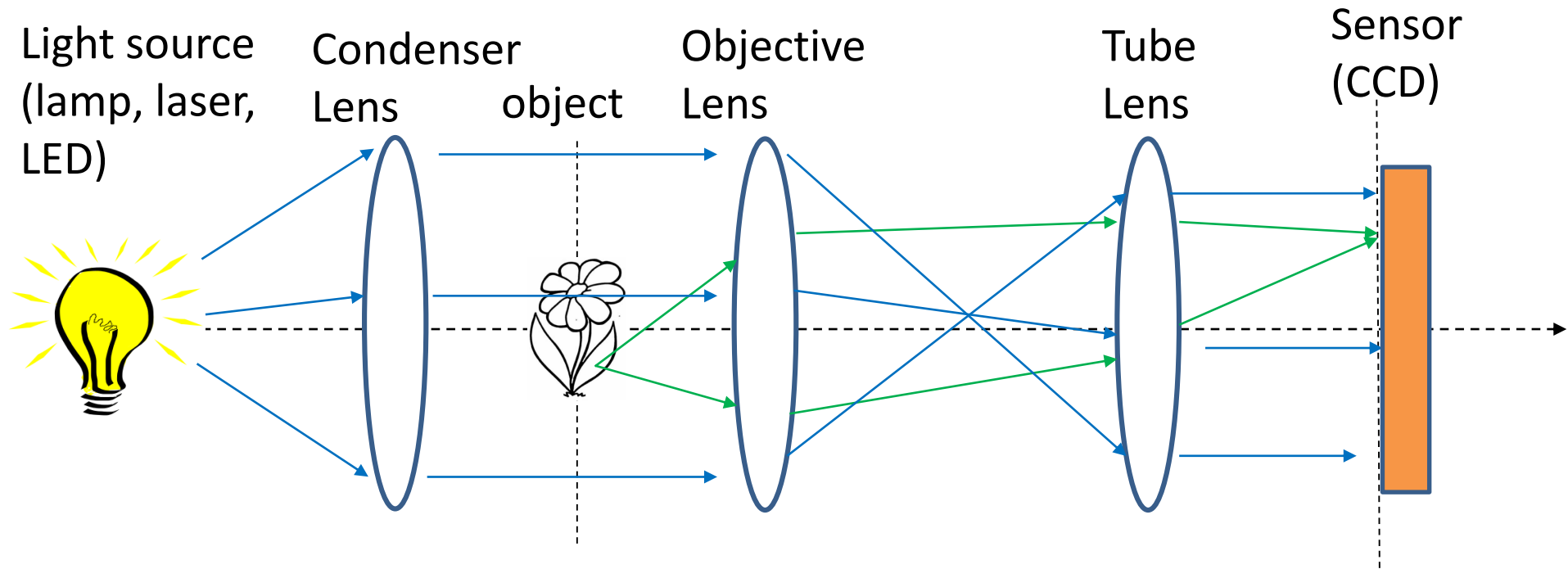


# Fluorescence Microscopy

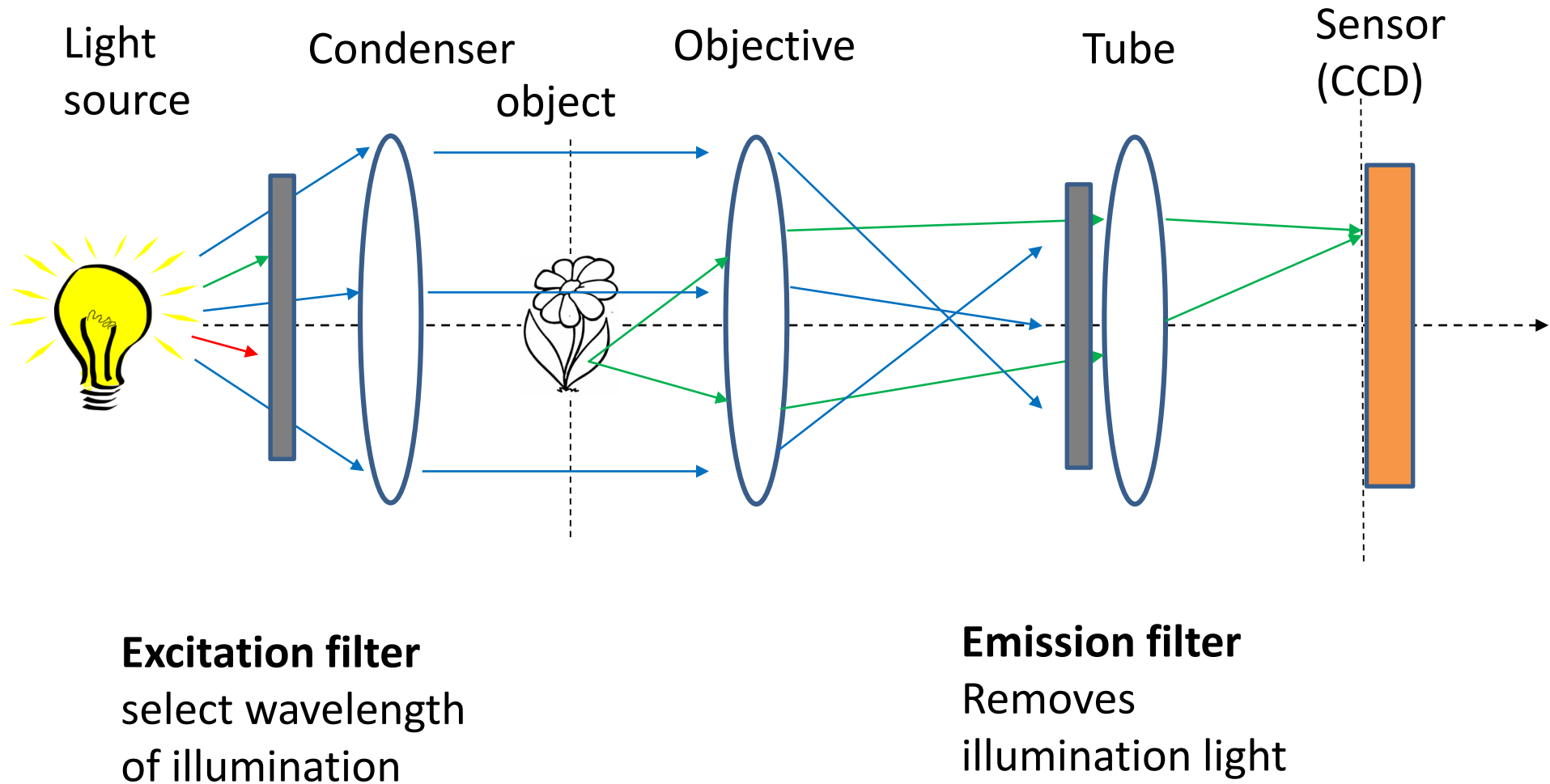
- Standard fluorescent microscope:
  - Easy & popular instrument
  - Relatively cheap (~25-50k)
  - Extremely versatile
  - Can be automated



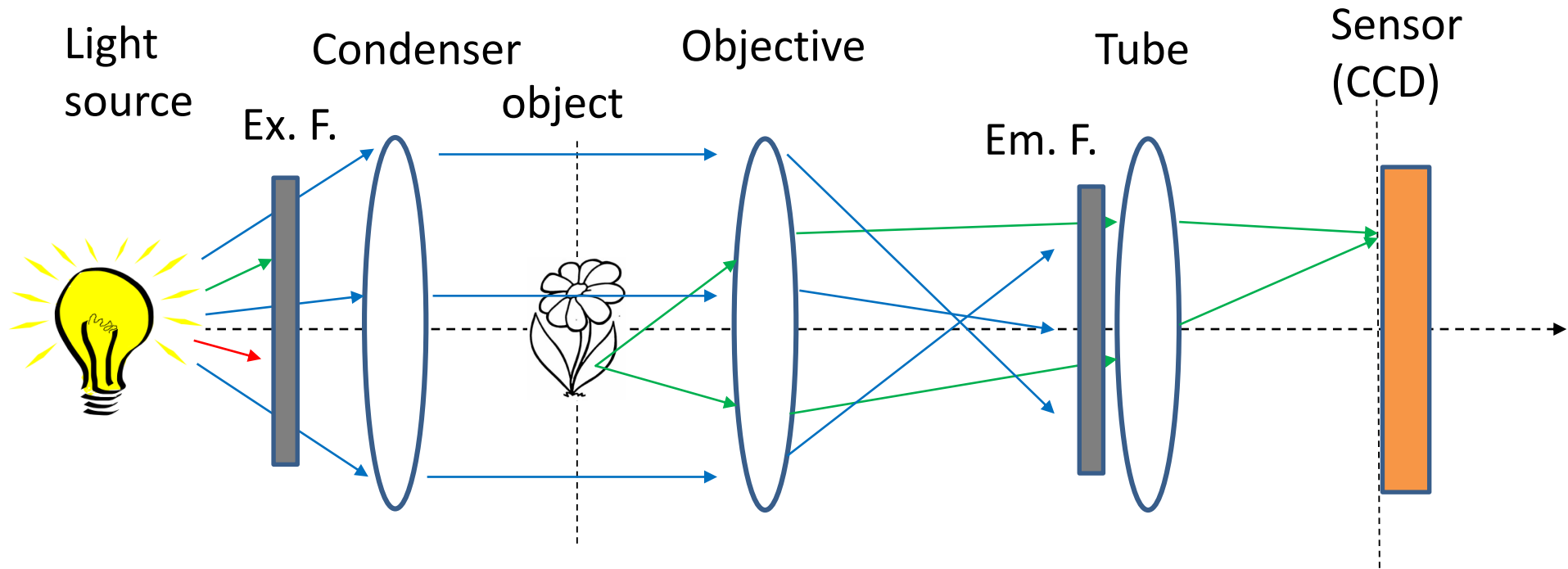
# Fluorescence Microscopy



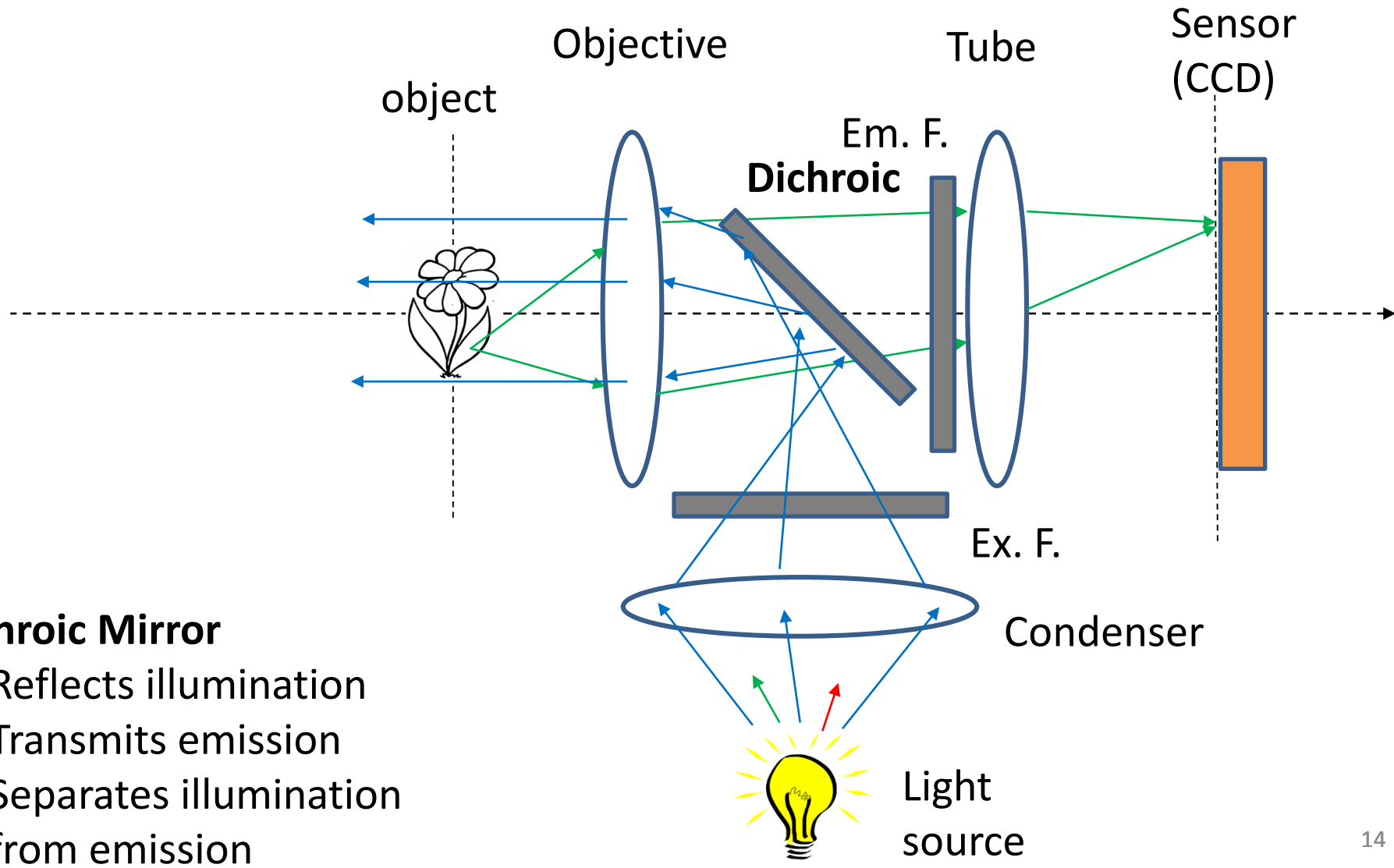
# Fluorescence Microscopy



# Fluorescence Microscopy



# Fluorescence Microscopy

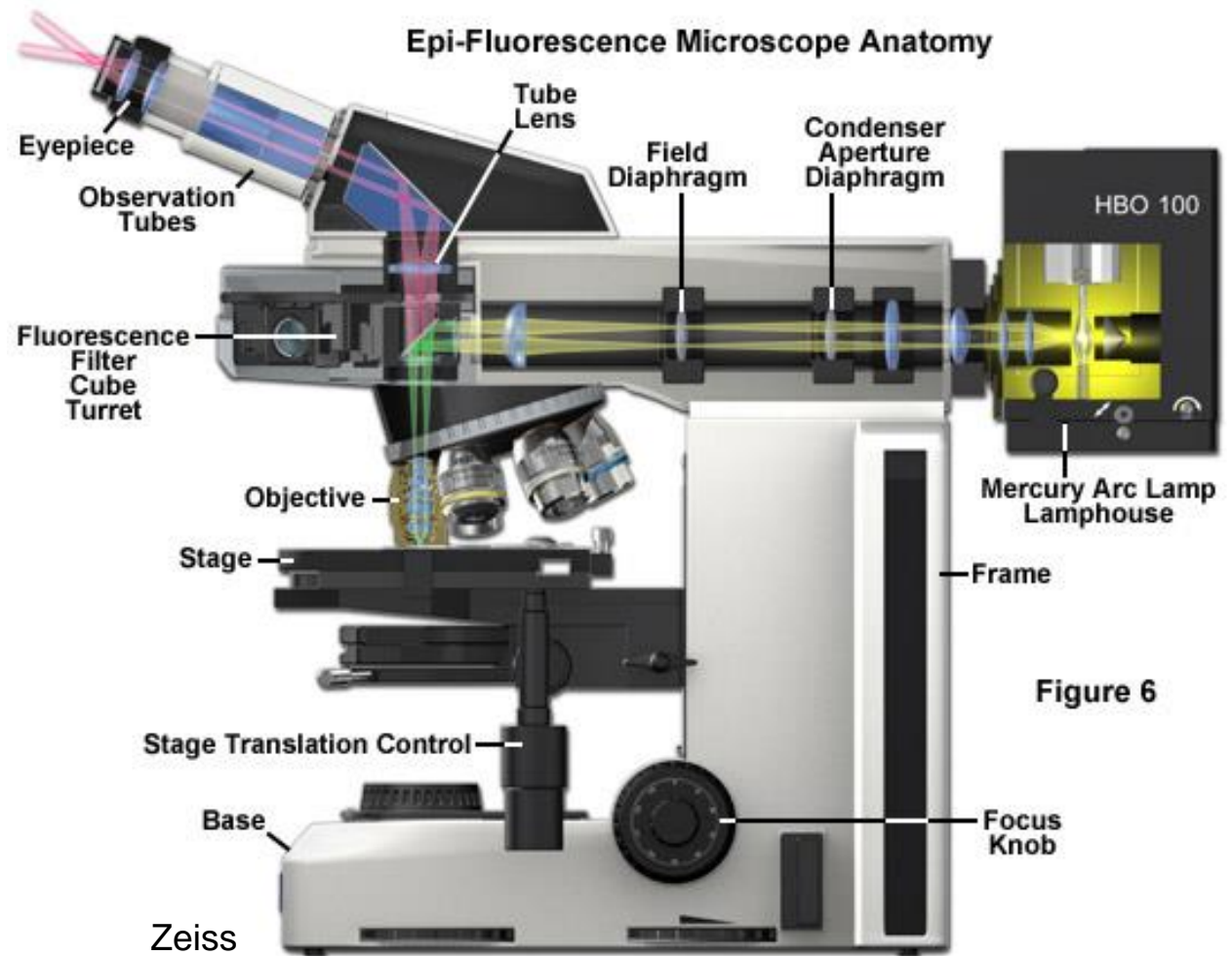


## Dichroic Mirror

- Reflects illumination
- Transmits emission
- Separates illumination from emission

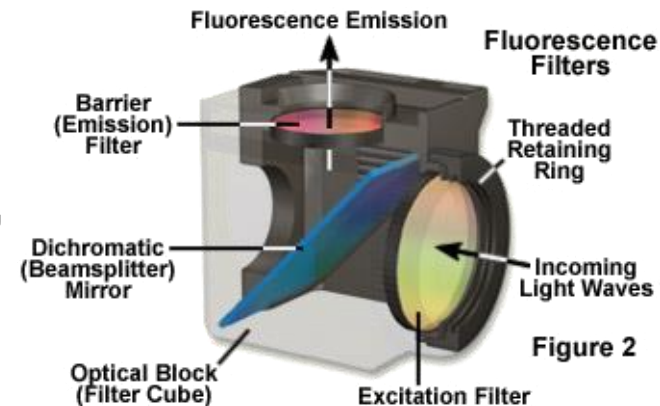
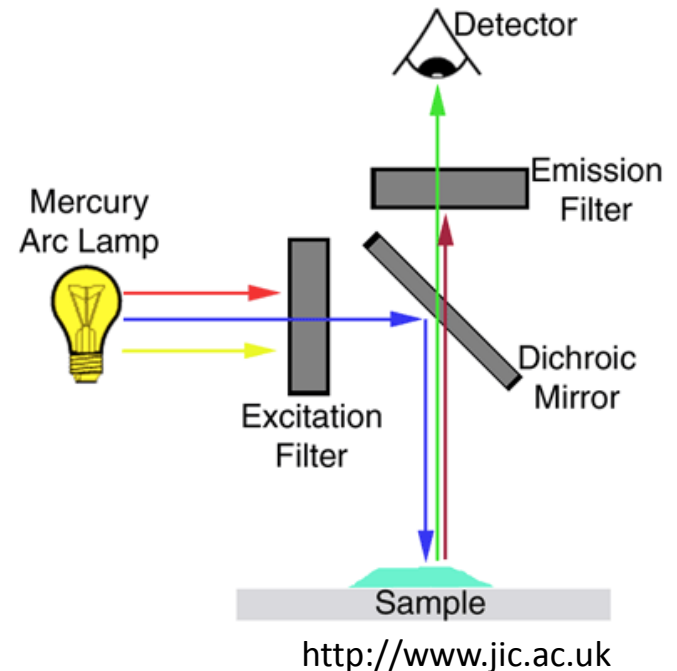
# Epifluorescence Microscopy

- The objective lens acts also as condenser for the illumination



# Fluorescence Microscopy Principle

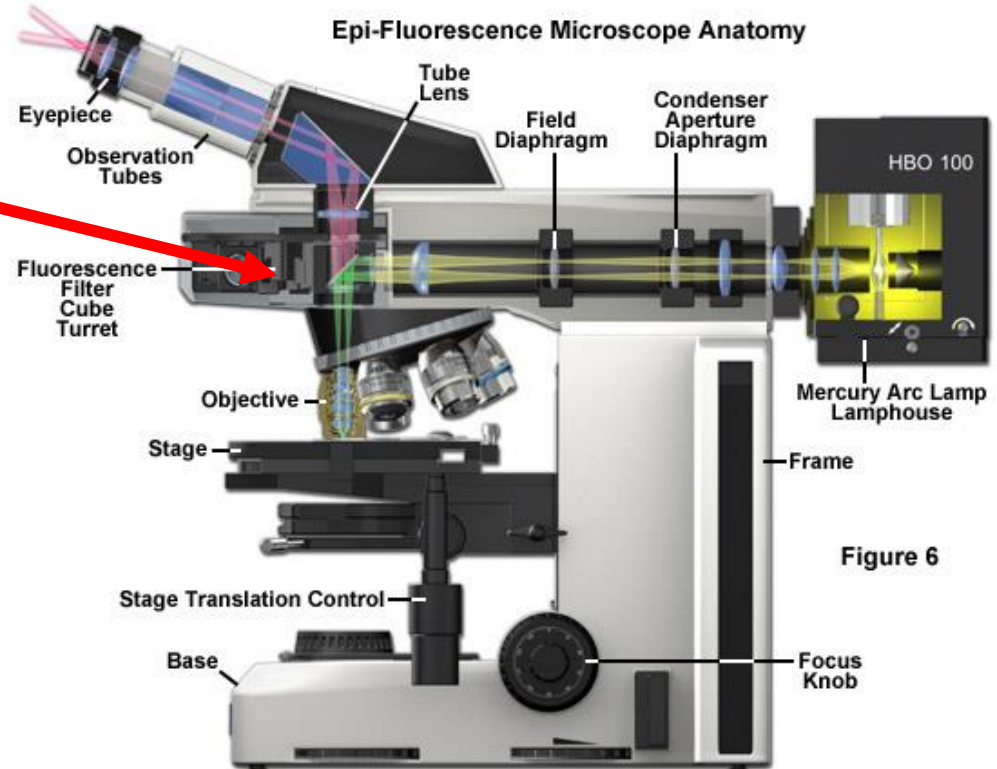
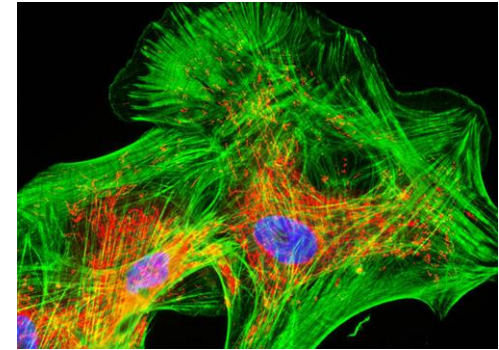
- A set of 3 optical filters is necessary for fluorescence microscopy
  - Excitation filter: selects wavelength of illumination
  - Dichroic filter: separates excitation from emission
  - Emission filter: removes remaining excitation light in emission path
- 3 filters placed in a “filter cube”
  - Need different filter set for each fluorophore color





# Filter Cubes

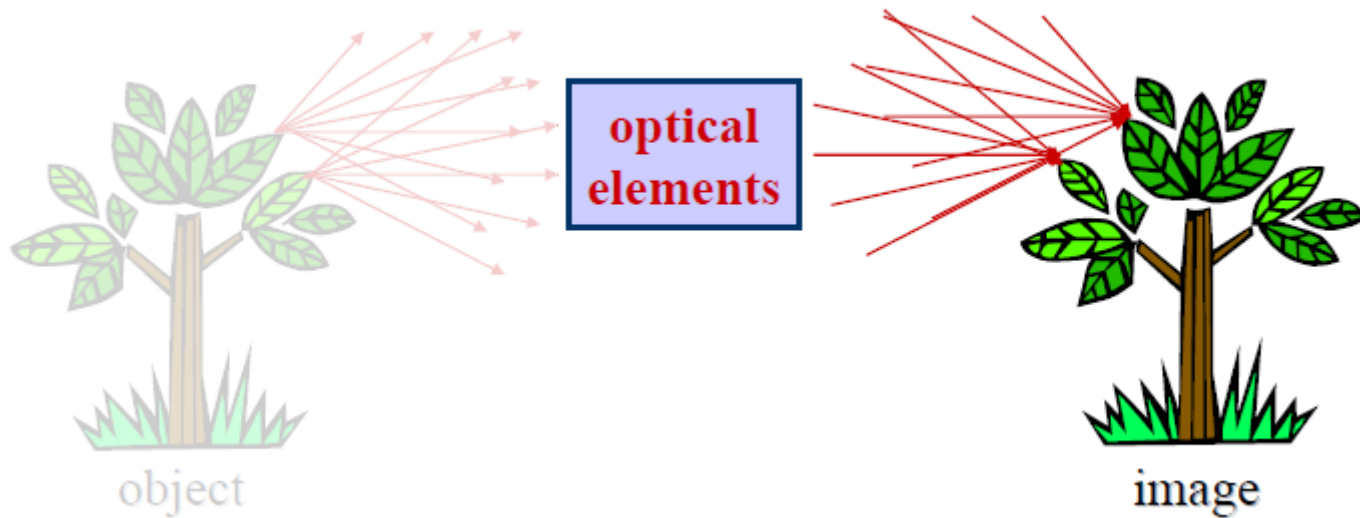
- Housed in a filter turret
  - Pick a different cube for each fluorophore color



# Fluorescence Microscopy Use

- Most modern biology/medical labs possess at least one
  - Ability to visualize expression & localization of any protein of interest
- Multi-color imaging
  - Need multiple “filter cubes”, one for each color

# State-of-the-Art in Fluorescence



# Live Cell Imaging

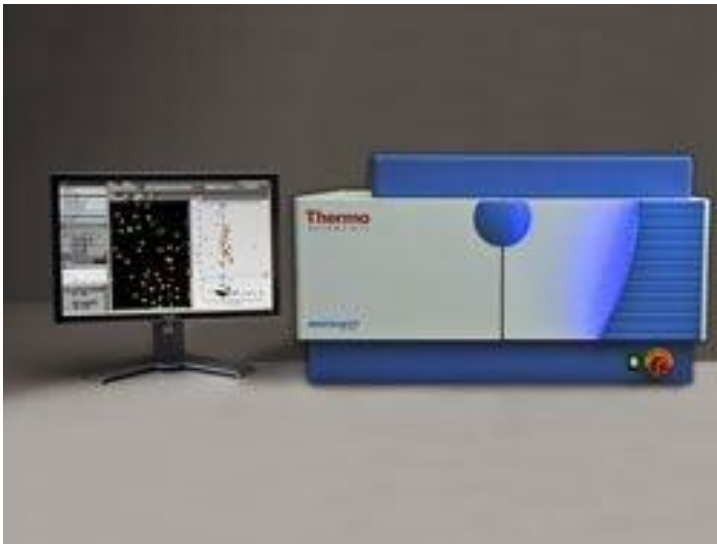
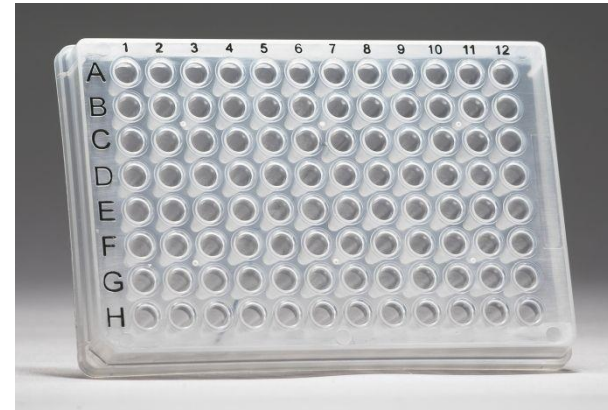
- Enclose microscope in a closet
  - Control temperature, humidity, CO<sub>2</sub> content → turn closet into a cell incubator
  - Image live cells as they proliferate/act



Olympus IX81 microscope

# Robotic Microscope

- Robotic microscopes that
  - Incubate cells in 96-well plates
  - Image sample over time
  - Able to image thousands of cell samples
  - Expensive (>150k)

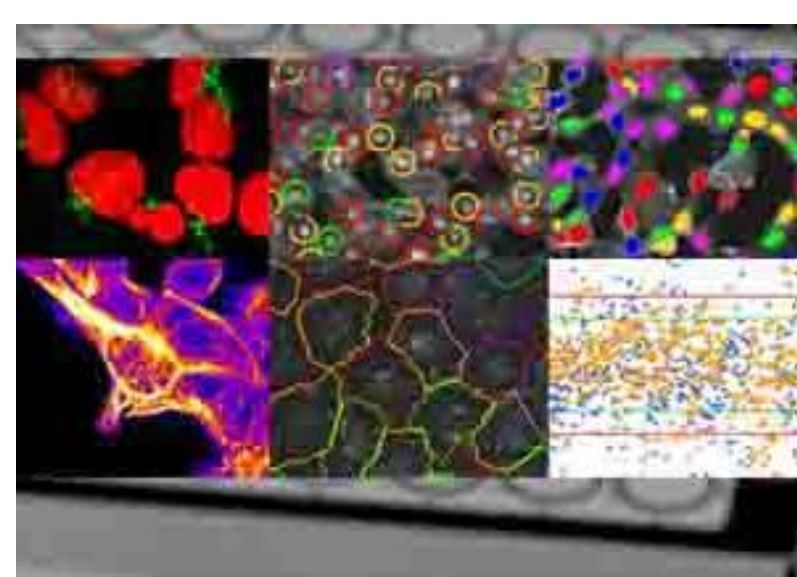


Thermo scientific arrayscan platform

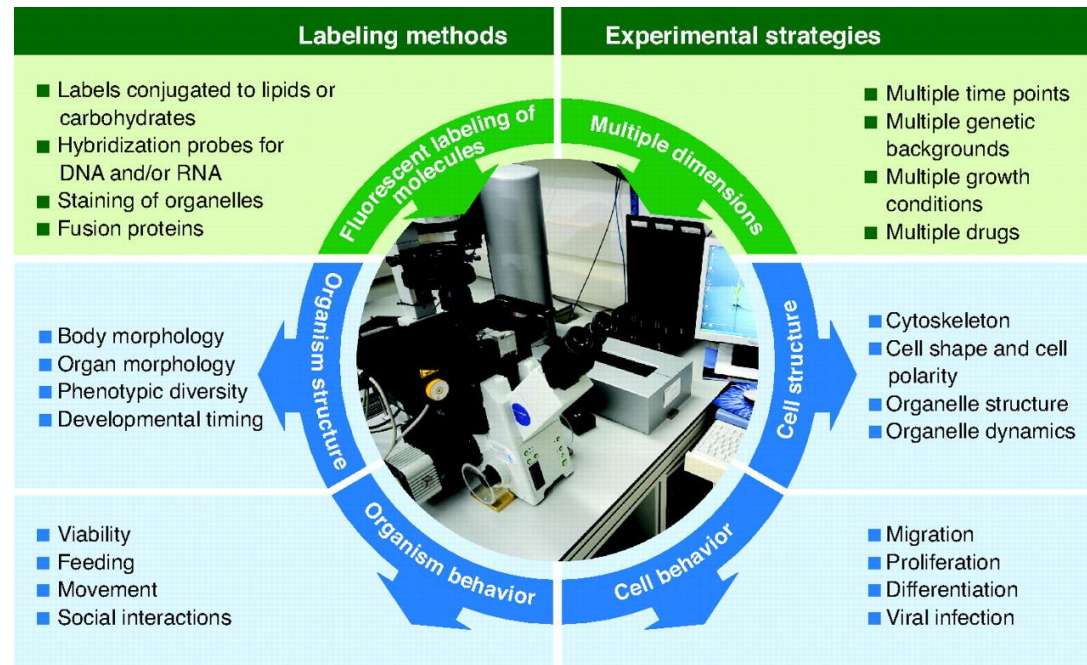


# High-Content Screening (HCS)

- Image multiple fluorophores in thousands of samples using robotics microscopes
- Computational tools analyze huge imaging datasets
- Used in drug discovery



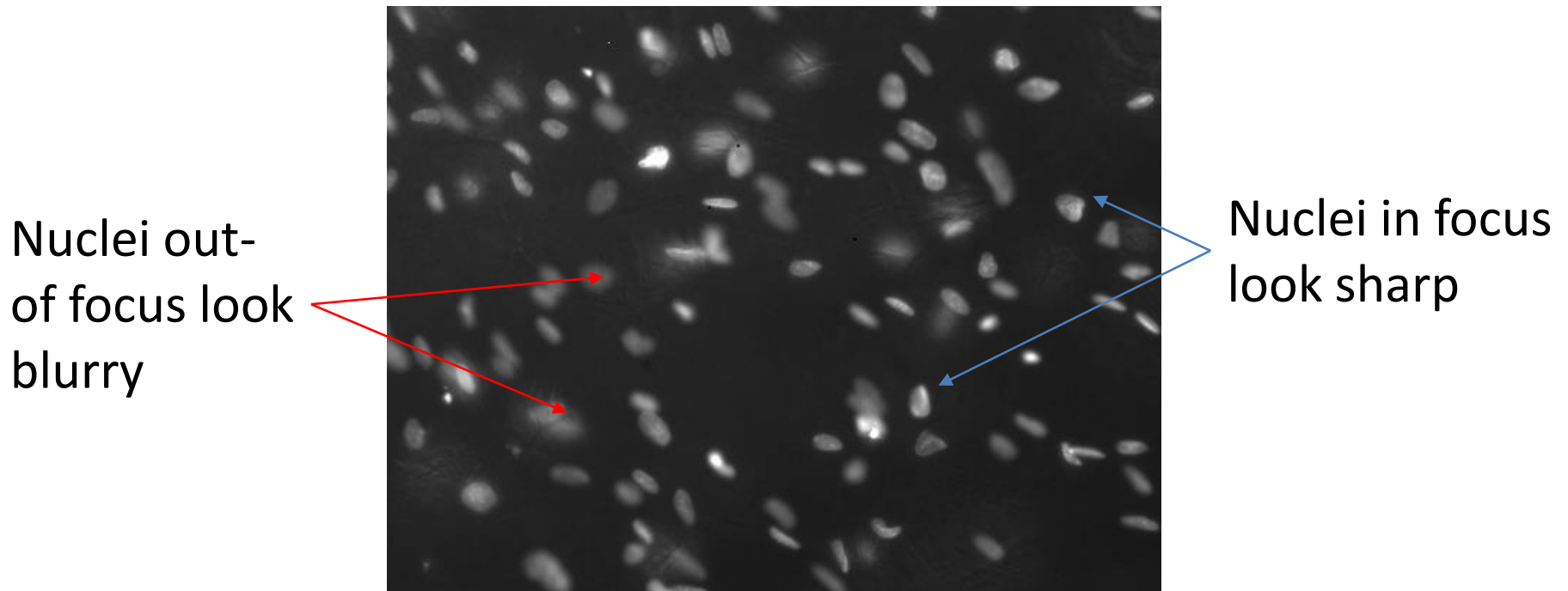
Assay development for HCS  
(Fraunhofer.de)



<http://jcs.biologists.org/content/124/22/3743>

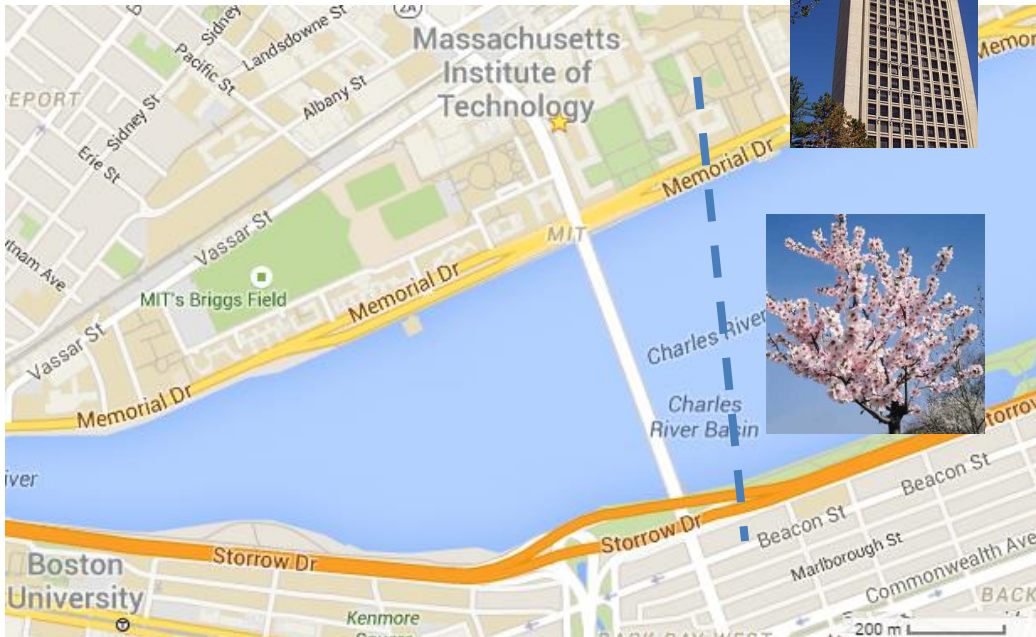
# Need to Overcome the Major Limitation of EpiFluorescence Microscopy

- Epifluorescence microscopy is a 2D imaging method
  - Suffers from out-of-focus light
  - Provides sharp image only of thin specimen ( $<20\text{ }\mu\text{m}$ )



# 2D Imaging

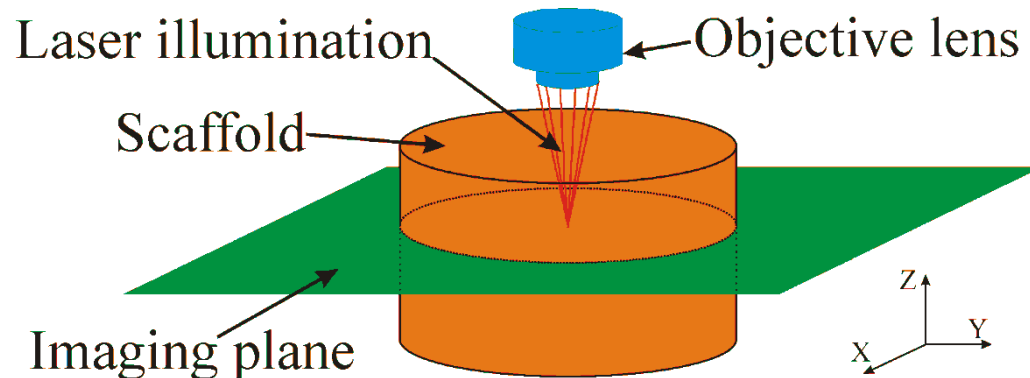
- In-focus objects imaged sharply
- Out-of-focus objects imaged blurry





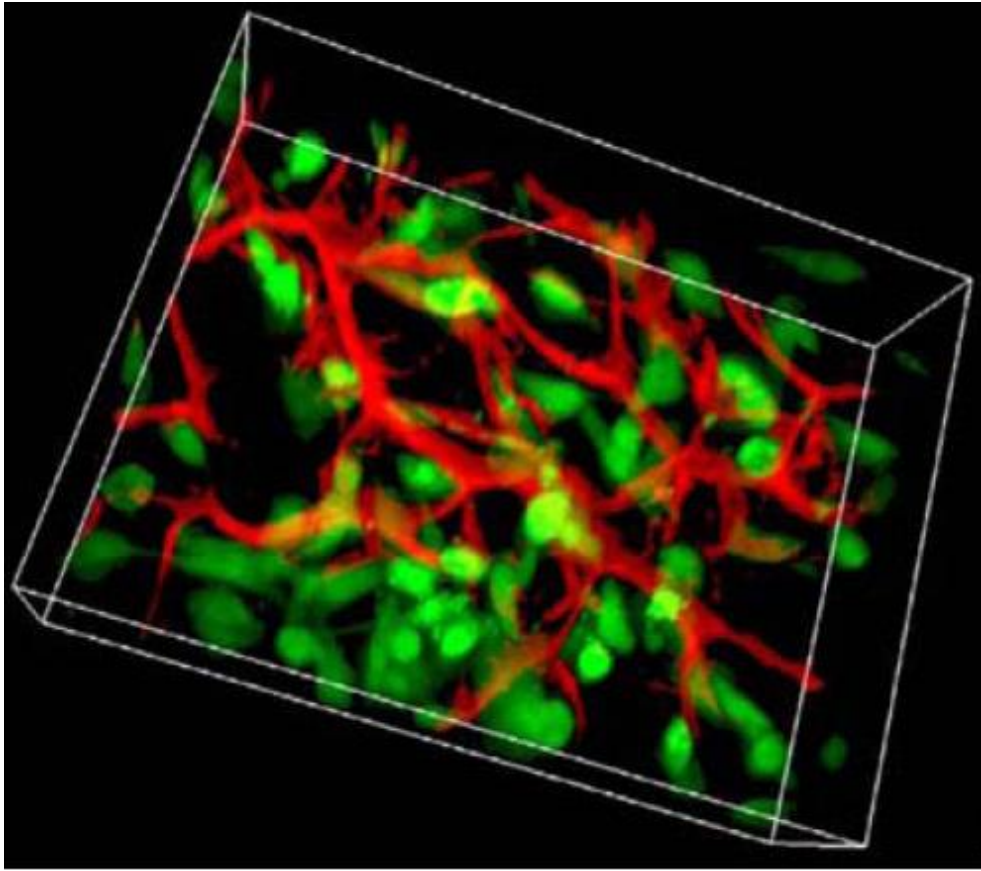
# 3D Imaging

- In-focus objects imaged sharply
- Out-of-focus objects are NOT imaged
- Image-slicing property
  - Image 1 thin ( $\sim 1\ \mu\text{m}$  thick) slice of the sample at any exposure



# 3D Imaging

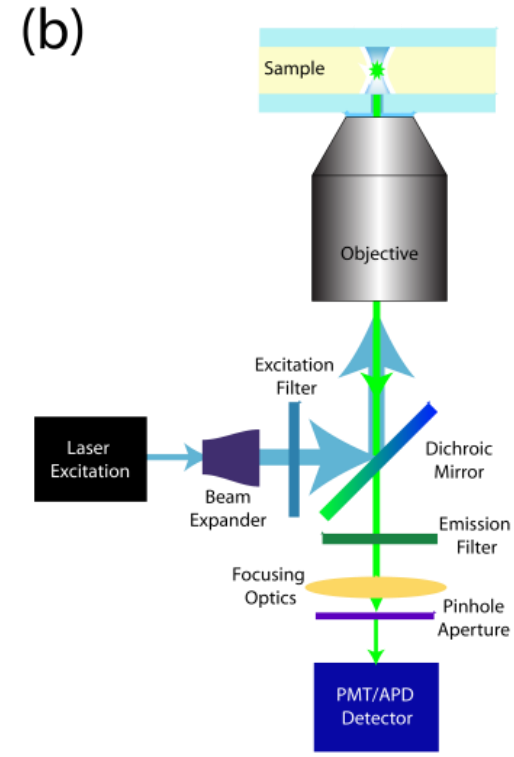
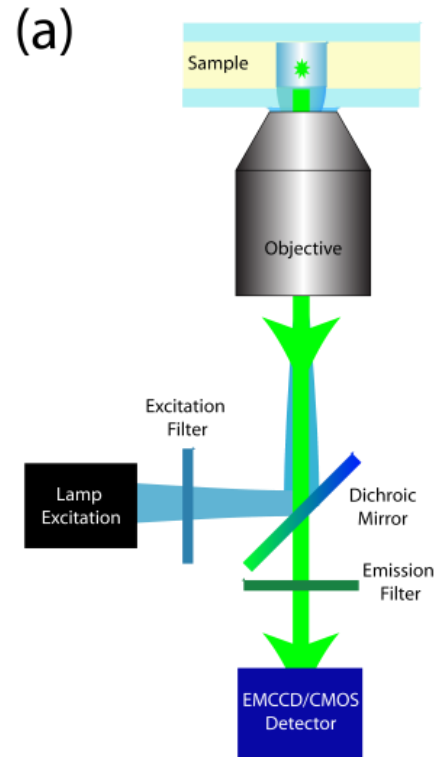
- A 3D image is generated by imaging various planes and combine them into a “image cube”



fibroblasts (green) inside a collagen scaffold (red). Harley, 2006

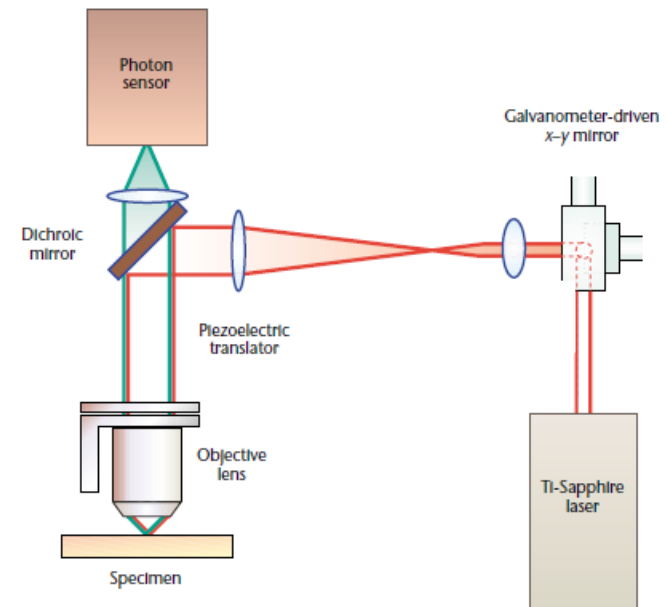
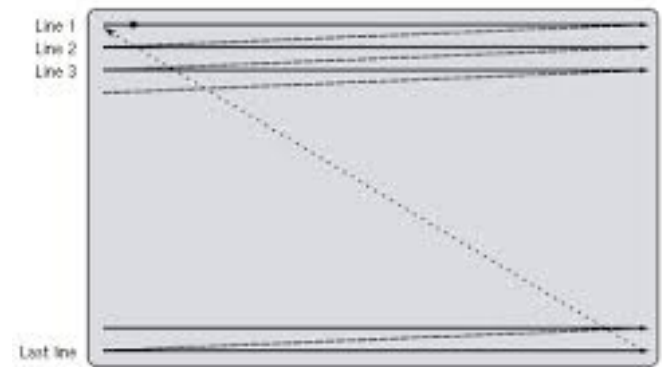
# Confocal Microscopy

- Optical Slicing is achieved in 2 ways
  - Focus illumination in the sample
  - Place pinhole before detector → reject out-of-focus light
- Use 1 pixel detector
  - PMT, photodiodes



# Confocal Microscopy

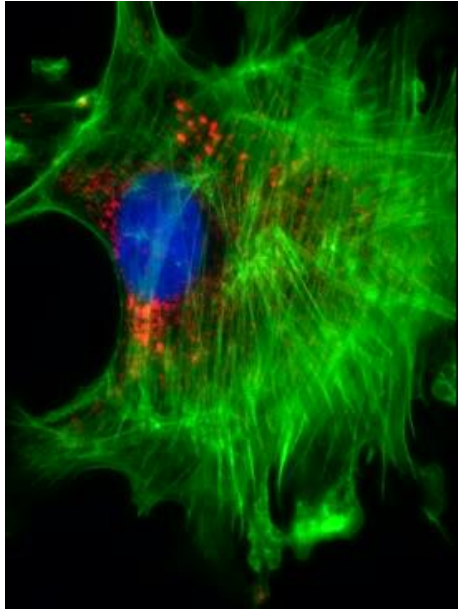
- At each time, a single photon is imaged
- Use scanning mirrors to control (x,y) location of laser focus
  - raster-scanning to image a plane
- Use piezoelectric actuators to shift objective
  - Sample along Z → image different planes



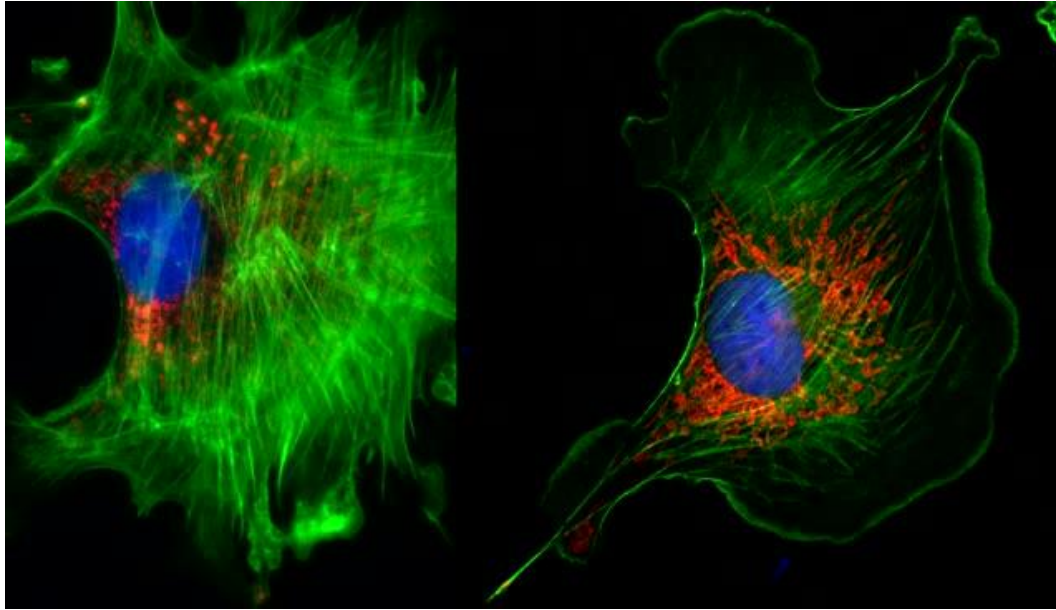
# Confocal Microscopy

- Most popular 3D imaging method
- Cost ~30-50k

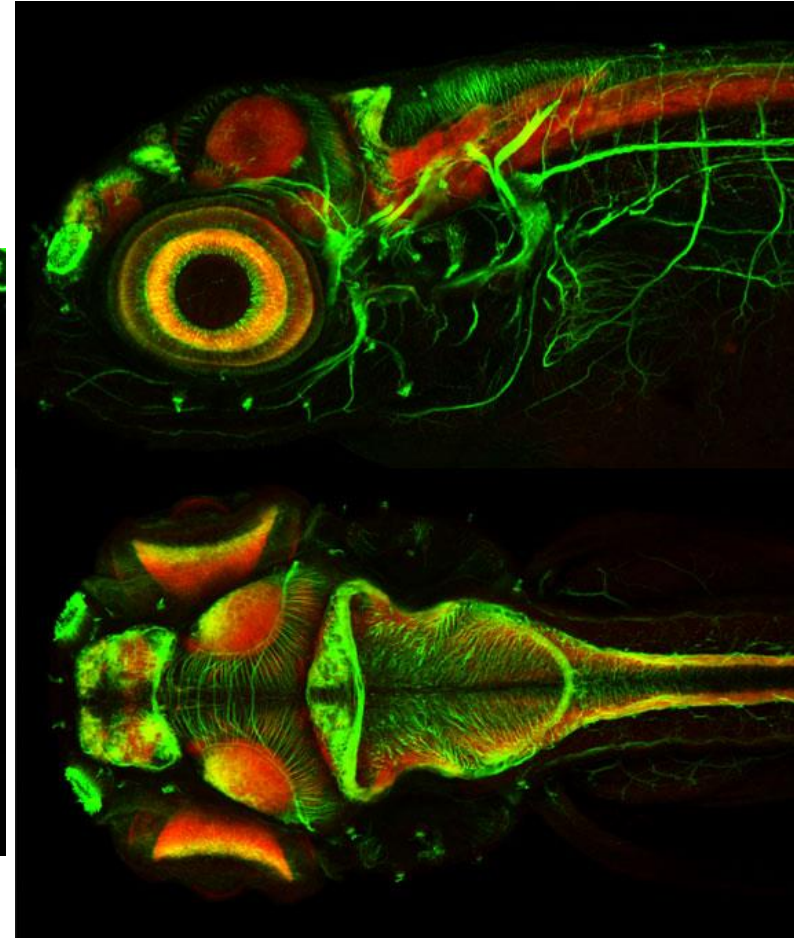
Epifluorescence



Confocal Imaging



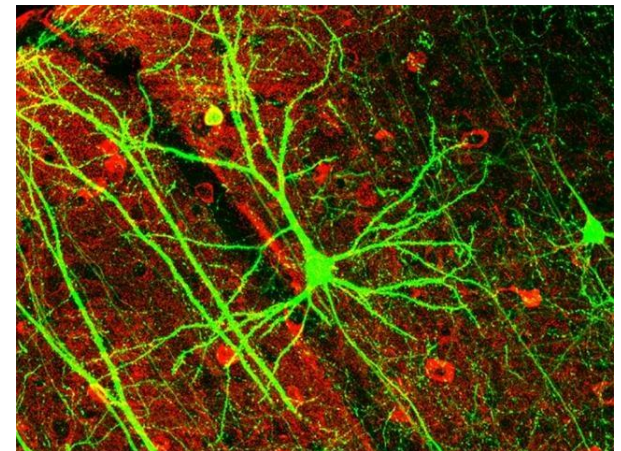
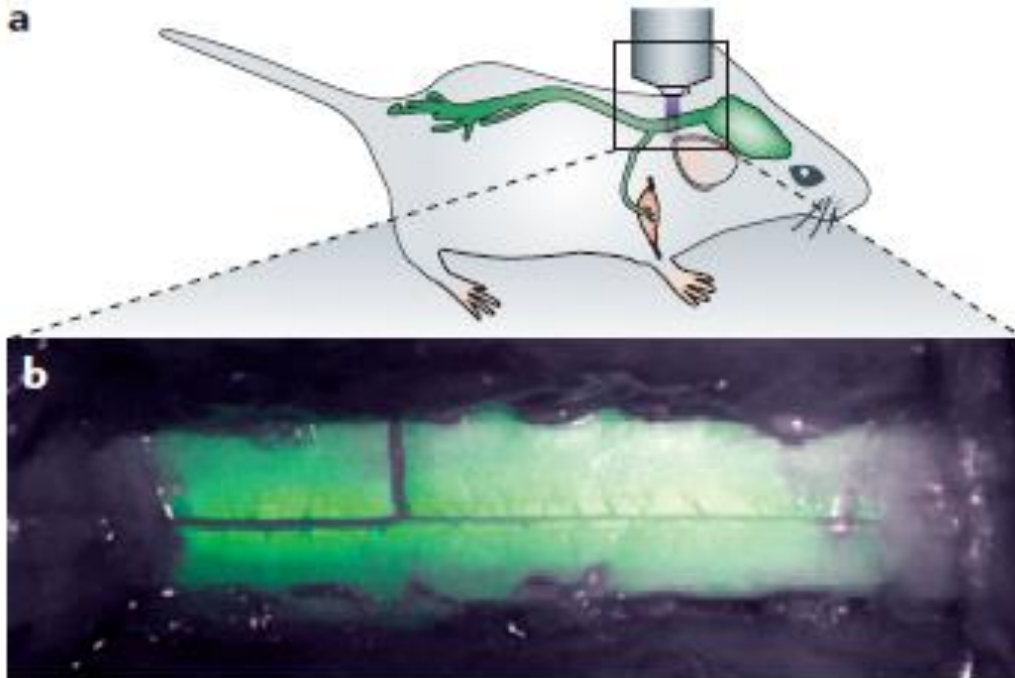
microscopysolutions.ca



4 day old zebrafish embryo labelled with SV2 and acetylated tubulin antibodies showing axon tracts(green) and neuropil(red). (<http://www.ucl.ac.uk/>)<sup>29</sup>

# *In vivo* imaging

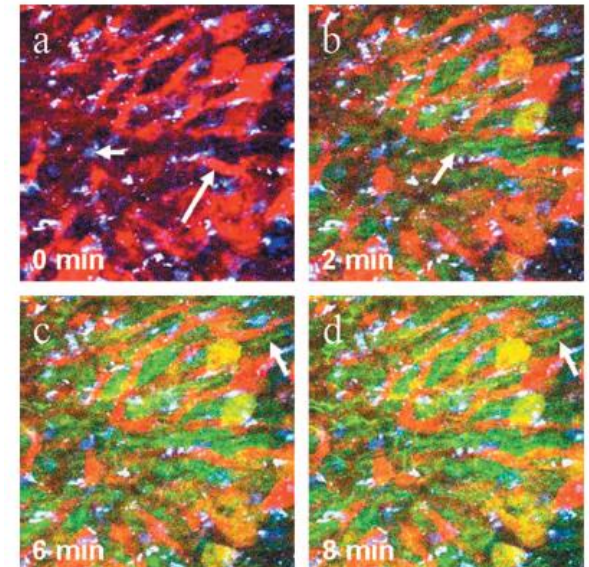
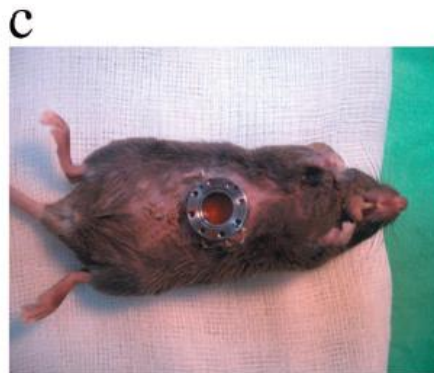
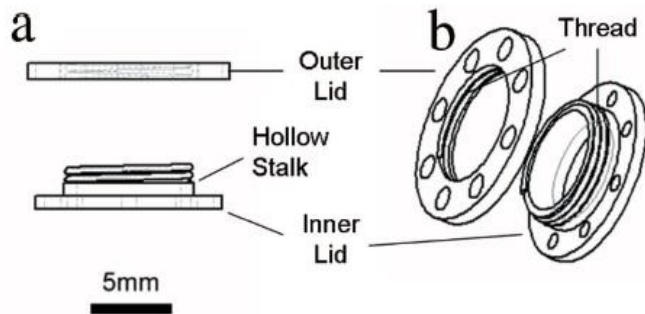
- Advanced 3D imaging methods (nonlinear optics) utilize IR light and provide 3D imaging
  - Enable 3D imaging in live transgenic animals
  - Applications: wound healing, neuron plasticity





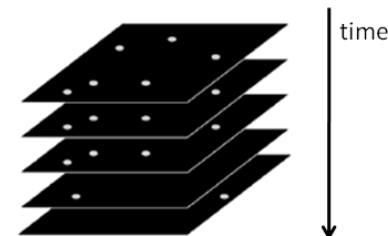
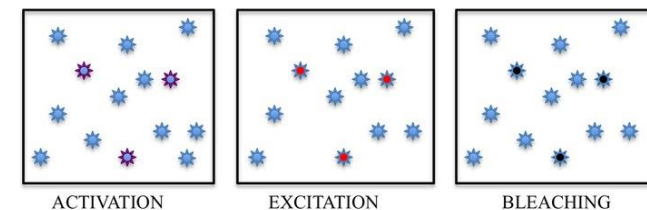
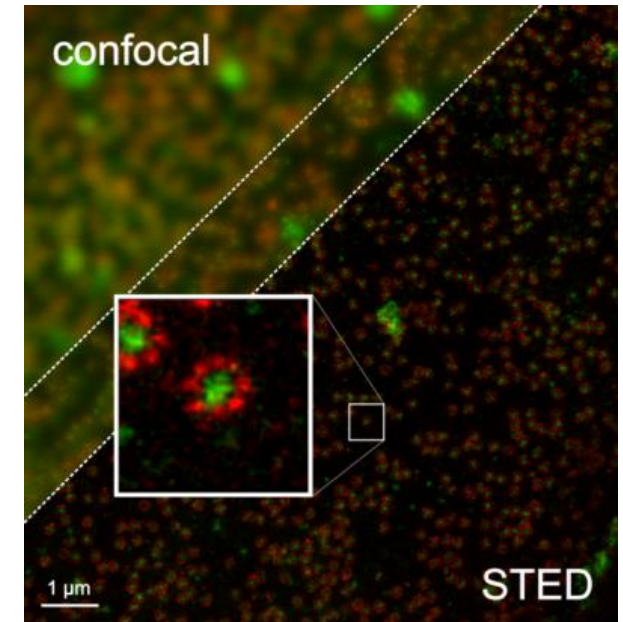
# *In vivo* imaging

- Engineers and doctors collaborate to develop optical devices that enable tissue imaging
  - Imaging cancer biology in animal models



# Super-Resolution Microscopy

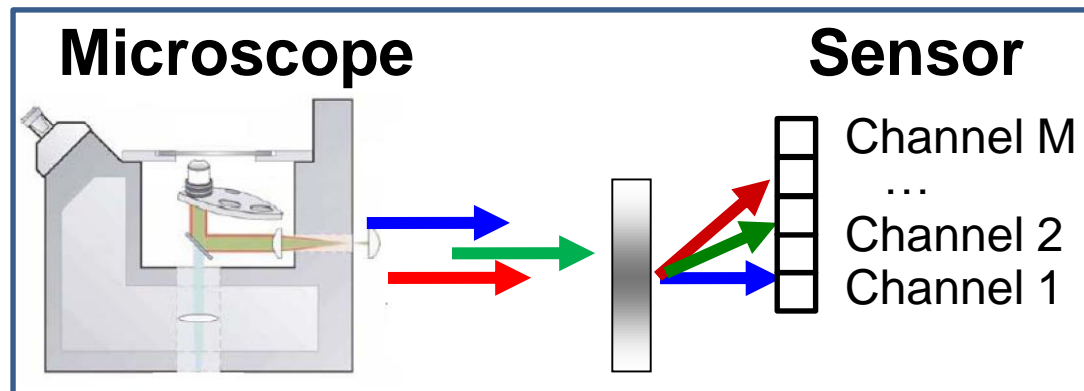
- Methods that provide optical resolution less than  $\delta r = 0.61 \frac{\lambda}{NA}$
- Methods:
  - STED, PALM/STORM
- PALM/STORM microscopy
  - Use photo-activated fluorophores
  - Light activate a small fraction of fluorophores  $\rightarrow$  image a sub-population of emitters at each exposure  $\rightarrow$  repeat many times



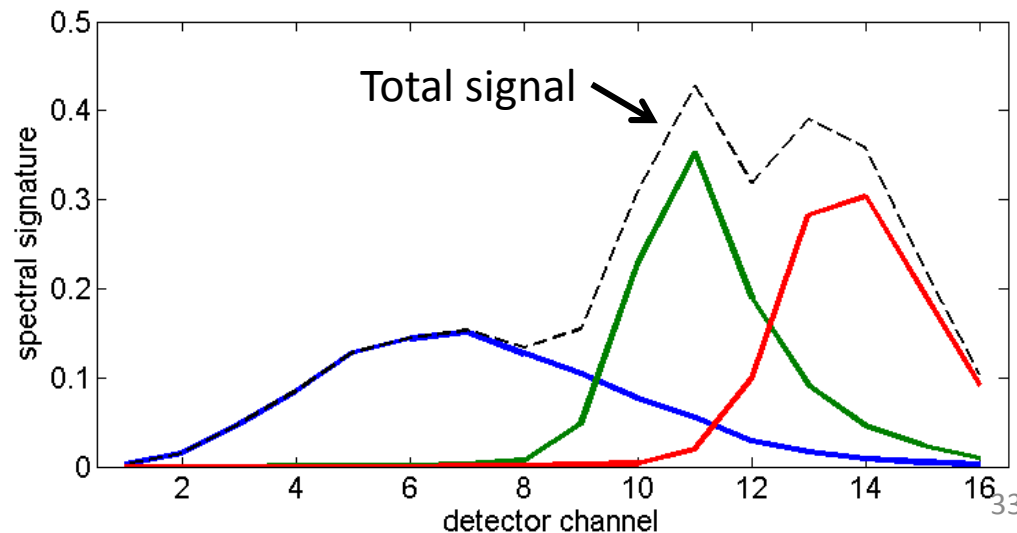


# Spectral Microscopy

- Resolve emission of multiple fluorophores based on their emission spectrum
- Use multi-channel detection systems
  - each exposure measures signal at several EM bands
- Use computation
  - Spectral unmixing

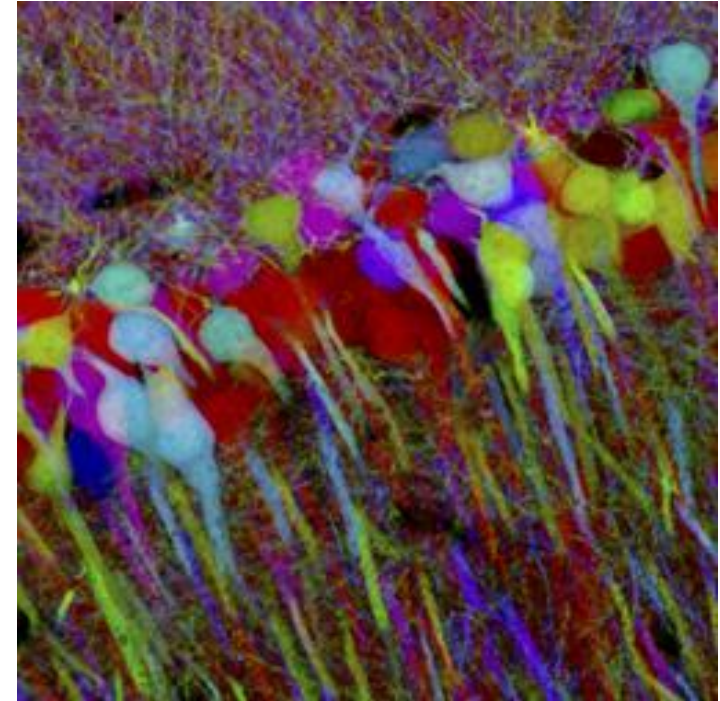
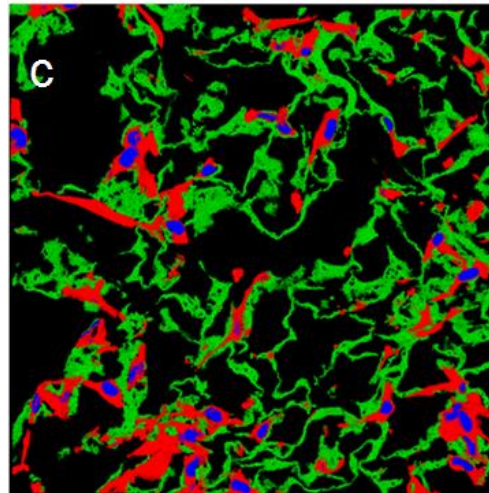
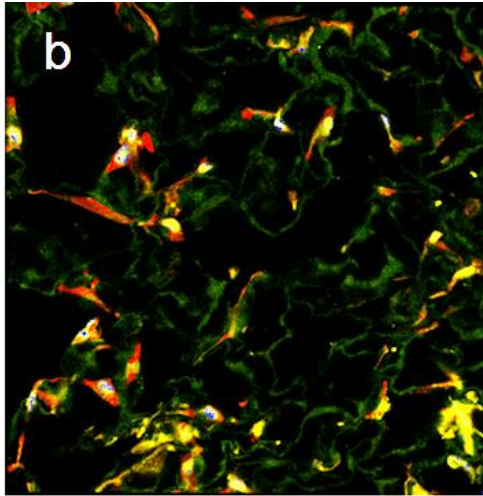


*Buelher et al. 2005*



# Spectral Microscopy

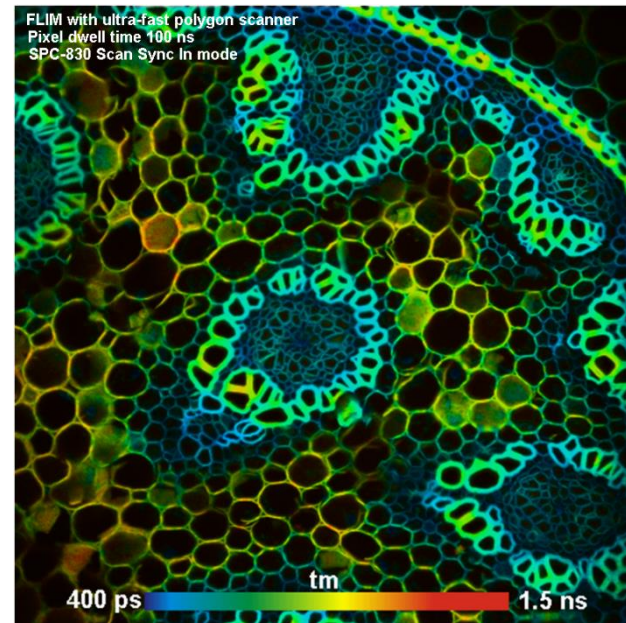
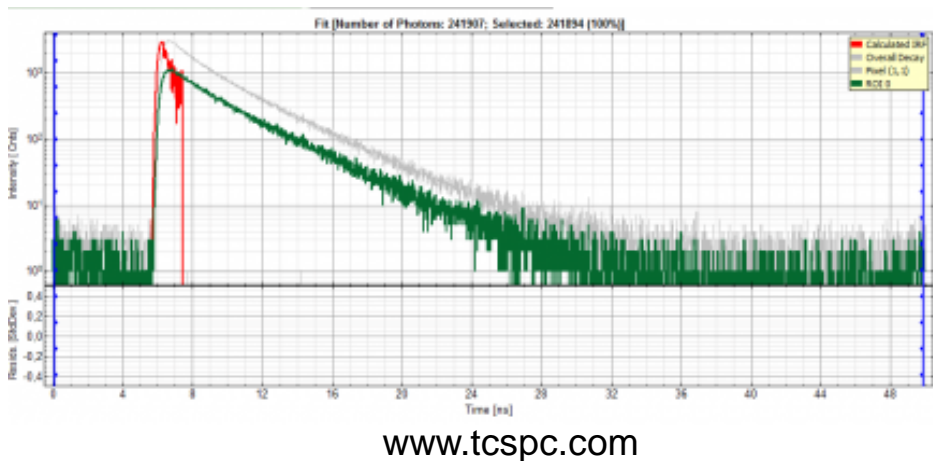
- Based on a single multi-channel image, quantify multiple emitters
  - Accelerate imaging
  - High content



■ second harmonic  
■ hoechst33342  
■ collagen  
■ alexa fluor 488  
■ CMTMR

# Fluorescence Lifetime Microscopy

- Instruments that can quantify the fluorescence lifetime at each pixel
- Exploit known fluorescence lifetime of each emitter to resolve the emission of each fluorophore



## Έκαναν το γλυκό πικρό μέσω της χειραγώγησης του εγκεφάλου

Επιστήμονες παρενέβησαν στον εγκέφαλο πειραματόζωνων και με την τεχνική της οπτογενετικής ενεργοποίησαν και απενεργοποίησαν συγκεκριμένους νευρώνες μέσω φωτός λέιζερ και άλλαξαν το είδος της γεύσης τους.

Δημοσίευση: 19/11/2015 - 15:55 Τελευταία ενημέρωση: 19/11/2015 - 15:55



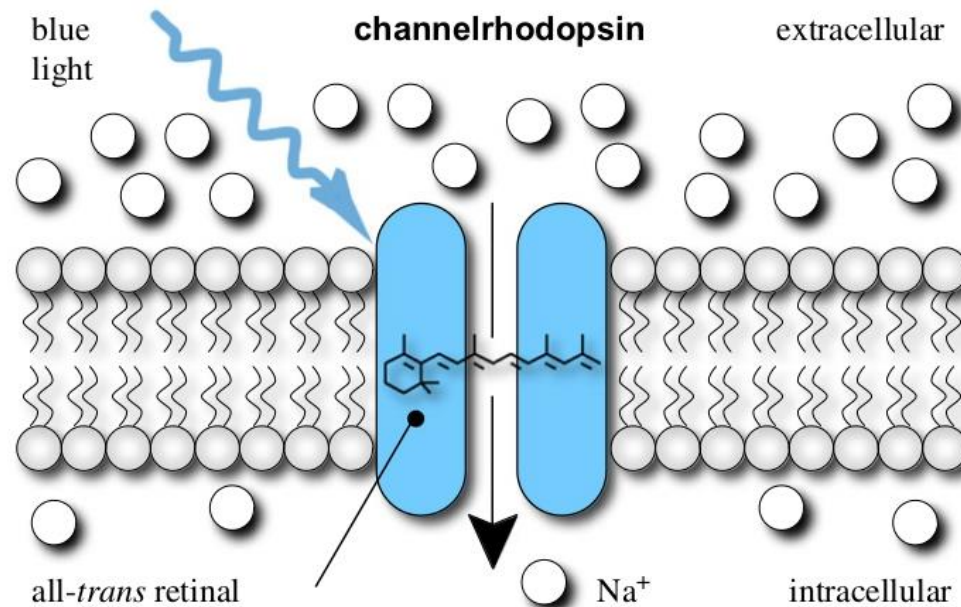
1 σχόλιο





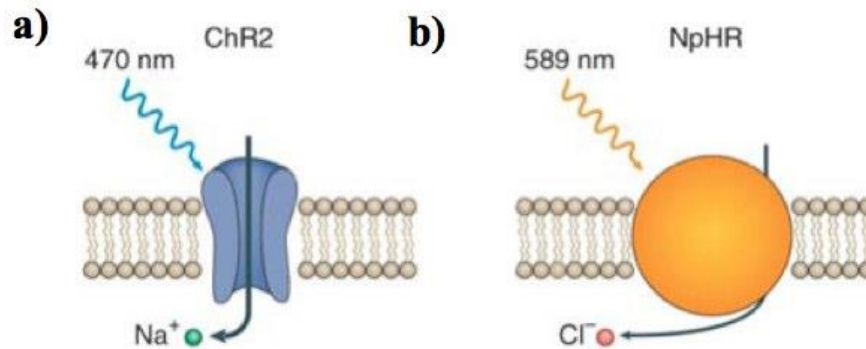
# Optogenetics

- Utilize light to activate/de-activate proteins
- Inspired by channelrhodopsin (chR)
  - Protein involved in imaging
  - Absorbs light → activated → opens a  $\text{Na}^+$  channel



# Optogenetics

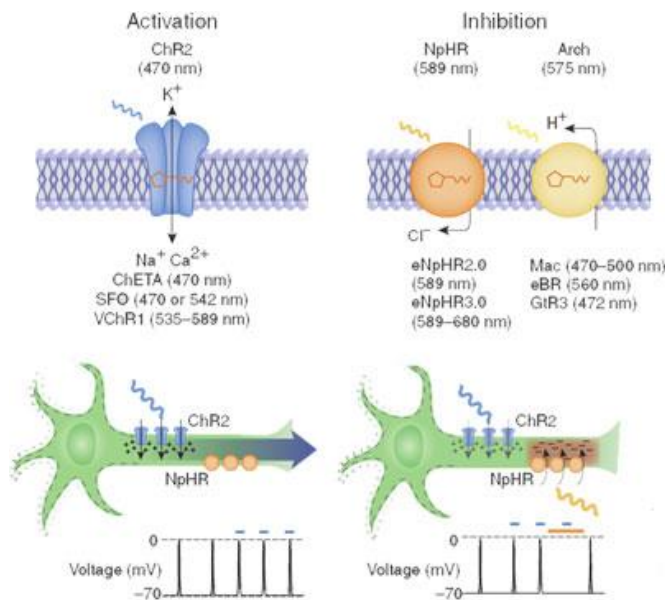
- Biological engineers modified chR → generated novel proteins that upon light activation can do other things
- Then transfect cells or generate transgenic animal that express these proteins



# Optogenetics



- Control/probe neuron circuits in brain → study brain
- Use microscope-like instruments to control light



Neuron activation/inhibition (nature)

